

The Role of Platelets in Tumour Growth

Úloha krevních destiček v rozvoji nádoru

Pilatova K.^{1,2}, Zdrzilova-Dubská L.^{1,2}, Klement G. L.^{2,3}

¹ Department of Laboratory Medicine, Masaryk Memorial Cancer Institute, Brno, Czech Republic

² Regional Centre for Applied Molecular Oncology, Masaryk Memorial Cancer Institute, Brno, Czech Republic

³ Center of Cancer Systems Biology, Steward St. Elizabeth's Medical Center, Pediatric Hematology Oncology, Tufts University School of Medicine, Boston, MA, USA

Summary

Platelets, as initial responders to vascular injury, play a very important role in the initial stages of the haemostatic process. While the role of platelets in coagulation has been well studied and documented, their role in other physiological and pathological processes is just emerging. Platelets contain many biologically active molecules and, as they adhere to sites of tumour activated or injured endothelium, many of these molecules are released into the local microenvironment leading to platelet-mediated effects on vascular tone, repair and neo-angiogenesis. Platelets are likely play important roles in the tumour microenvironment that may be thought of as "a wound that never heals".

Key words

blood platelets – angiogenesis – wound healing – tumour growth – neoplasm metastasis

Souhrn

Krevní destičky jako elementy odpovídající v první vlně na poškození cév hrají velmi významnou úlohu v počátečních fázích procesu hemostázy. Zatímco zapojení trombocytů v procesu koagulace je podrobně studováno a popsáno, jejich role v dalších fyziologických a patologických procesech teprve začíná být předmětem zájmu. Krevní destičky obsahují řadu biologicky aktivních molekul a s tím, jak trombocyty adherují na nádorem aktivovaný nebo poškozený endotel, je řada těchto molekul uvolňována do nádorového mikroprostředí, což vede k ovlivnění cévního tonu, reparaci cévy a neoangiogenezi. Destičky pravděpodobně hrají důležitou úlohu v mikroprostředí nádoru, který můžeme považovat za ránu, která se nehojí.

Klíčová slova

trombocyt – angiogeneze – hojení rány – růst nádoru – metastazování nádoru

The work was supported by the European Regional Development Fund and the State Budget of the Czech Republic for Regional Centre for Applied Molecular Oncology (RECAMO, CZ.1.05/2.1.00/03.0101).

Práce byla podpořena Evropským fondem pro regionální rozvoj a státním rozpočtem České republiky (OP VaVpl – RECAMO, CZ.1.05/2.1.00/03.0101).

The authors declare they have no potential conflicts of interest concerning drugs, products, or services used in the study.

Autoři deklarují, že v souvislosti s předmětem studie nemají žádné komerční zájmy.

The Editorial Board declares that the manuscript met the ICMJE "uniform requirements" for biomedical papers.

Redakční rada potvrzuje, že rukopis práce splnil ICMJE kritéria pro publikace zaslané do biomedicínských časopisů.



Giannoula Lakka Klement, M.D.
Center of Cancer Systems Biology
Steward St. Elizabeth's Medical Center
Pediatric Hematology Oncology
Tufts University School of Medicine
Boston, MA, USA
e-mail: giannoula.klement@tufts.edu

Submitted/Obdrženo: 12. 11. 2012

Accepted/Přijato: 15. 11. 2012

The Hypercoagulable State Associated with Malignancy

Numerous clinical and basic science studies corroborate the importance of thrombosis in cancer development [1–7], cancer progression [8–12], and cancer metastasis [8,13–18]. The association is so well known that a deep vein thrombosis (DVT) in a patient without obvious risk factors triggers a search for an occult cancer. Despite this appreciation of a link between DVT and malignancy [19–21], the underlying biology has not been well characterised. The propensity to develop thromboembolic disease varies with the type of cancer [22], suggesting tumour cell-specific or tumour microenvironment-specific pathways to platelet and fibrin aggregation in tumours. Furthermore in some tumours, such as neuroblastoma, high platelet counts are associated with good prognosis [23], whereas in others (lung, colon, cervical, and breast cancers), the finding of high platelet counts implies poor prognosis [24–26].

Even though the association of hypercoagulability in cancer was first documented by Trousseau in 1865 [27], much work remains before we can use this finding therapeutically. There are some encouraging clinical observations. For example, the use of anticoagulants provides cancer patients with a survival advantage over and above that which would be conferred by the treatment of the DVT alone [28–35]. Unfortunately, large studies of the use of anticoagulants in the cancer population have not led to any significant change in the present management of cancer patients [29,36]. Yet both clinicians and basic scientists appreciate that even in patients not presenting with a cancer-associated thrombosis, the coagulation system is activated and platelet turnover increased. The interplay between platelets, coagulation and cancer is yet to be fully explored.

The Role of Platelet in Angiogenesis

The first scientific evidence suggesting that platelets were necessary for vascular integrity was reported in the late 1960's [37]. Organs perfused with pla-

telet poor plasma led to loss of integrity of the endothelial cell layer and haemorrhages, and this effect could be reversed by addition of platelets. Similarly, thrombocytopenia was associated with increase in vascular permeability due to large endothelial wall fenestrations (EC) [38,39]. Based on these and other studies platelets were thought to promote endothelial cell growth [40], even though the mechanism of this trophic effect was unclear.

Platelets contain three types of granules: α -granules, dense granules and lysosomes, but most angiogenesis related proteins are contained in α -granules [41,42]. Tab. 1 lists angiogenesis regulators found in platelets. The presence of proteins with opposing angiogenic functions in platelets suggests that platelets are mediators and their presence can result in different actions depending on the situation. The formation of a clot not only provides a matrix facilitating cell migration, but also leads to a very judicious release of either stimulators or inhibitors of growth. As platelets adhere to activated endothelia or to exposed vascular sub-endothelia, the reciprocal interactions between the cells lead to sequential release of angiogenesis regulators. Platelets in this way serve as potent activators as well as inhibitors of important tissue repair processes such as inflammation [43] and angiogenesis [44].

Platelets in Tumour Angiogenesis

A tumour is a community of cells. There are resident cells (fibroblasts, histiocytes, epithelial and mesenchymal cells) that form the tissues, and cells that are recruited to the site in time of injury or malignant growth (mesenchymal progenitors and inflammatory cells). Platelets are mediators of this community.

Primary tumour growth is facilitated by inflammation and angiogenesis not unlike physiological wound healing [86–88]. However, in cancer, the normal physiological processes of dialing-down angiogenesis as scar tissue develops is prevented by the continuous, onco-gene-mediated induction of tumour angiogenesis [89–92]. It has been well

described that tumour vasculature is immature, unstable and morphologically different from the normal systemic vasculature. While tumour vasculature is often thought of as abnormal, it is better conceptualised as an unpruned, underdeveloped precursor of mature vessels – a continuously expanding, but not maturing, vascular bed.

Platelets play an important role in modulating tumour dynamics. A large body of evidence spanning at least four decades supports the involvement of platelets in cancer [1,2,13]. The process of sequestration of angiogenesis regulators in platelets is an active and highly selective process [41]. An open-ended proteomic comparison of platelets from mice bearing dormant or fast-growing liposarcoma xenografts revealed significant differences in protein profiles between each of these tumour subtypes [41,93,94], as well as differences when platelets of mice bearing either of the tumour types were compared to platelets of non-tumour-bearing sham-operated controls. Despite the open-ended analysis of all proteins present in platelets, the majority of proteins differentially expressed in platelets of tumour-bearing animals and cancer patients were found to be angiogenesis regulators such as VEGF, bFGF, PDGF, PF4, TSP1, MMP9, endostatin, angiopoietin-1 and -2, etc. While the membership in this “platelet angiogenesis proteome”, as well as the concentrations of individual protein members, is fairly stable under physiological conditions [45], it is altered very early in tumour growth [41,93]. The sequestration of angiogenesis regulators in platelets is: i) *active* because it occurs against a concentration gradient in plasma and ii) *highly selective* for angiogenesis regulators, as other very abundant proteins, e.g. albumin or fibrinogen, are not taken up by platelets against a concentration gradient. Interestingly, the sequestration of angiogenesis regulators in platelets occurs very early in primary tumour growth [93]. At a time when tumours are not detectable by conventional methods, and long before the tumour burden results in changes in the levels of angiogenesis regulators in plasma or serum, there are

Tab. 1. Summary of pro- and antiangiogenic actions of platelet cytokines and factors.

ANGIOGENESIS STIMULATORS			
Factor	Platelet concentration	Mechanism of action	Ref.
VEGF (all isoforms, mainly VEGF-A, -B)	0.74 ± 0.37 pg/10 ⁶ PLT	Promotes permeability of the vessel wall and serves as a chemoattractant for EC sprouting in the initial stage of the angiogenic response.	[41,45,46]
PDGF	23 ± 6 pg/10 ⁶ PLT	Stimulates proliferation, differentiation and migration of fibroblasts or smooth muscle cells, providing support for the newly formed angiogenic sprout in the form of the pericyte (advanced stages of the angiogenic process).	[45,47,48]
FGFs (aFGF, bFGF/FGF-2)	bFGF: 0.44 ± 0.15 pg/10 ⁶ PLT	Serve as a chemoattractant for EC and stimulate proliferation of EC.	[45,49]
EGF	1.05 pg/10 ⁶ PLT	EGF binding to specific receptor EGFR induces an EC response leading to increased tubule formation, cell division and movement. EGF can augment the proangiogenic effect of other factors.	[50–52]
HGF	–	HGF is mitogen for different cell types including EC. Mechanisms of its effects include stimulating of secretion of MMP-1, VEGF, HGF itself and its receptor, c-met, in EC. Alternative processing of the HGF α-chain mRNA produces anti-angiogenic fragments.	[53–55]
IGF	–	Stimulates VEGF mRNA synthesis in EC. Facilitates EC motility and tubule formation.	[56–59]
angiopoetins	–	Ang-1 stimulates EC migration, tube formation, sprouting, and survival.	[60–61]
SDF-1/CXCL12	–	SDF-1α expression on activated platelet surface enhances endothelial progenitor cell recruitment to sites of arterial injury.	[62]
CD40L/CD154	–	CD40L binding to CD40 on EC promotes EC proliferation, migration and vessel-like structure formation through activation of the PI3K/Akt signalling pathway.	[63]
MMPs	–	Cleave different components of extracellular matrix (ECM) and basement membrane, which support new vessel development by assisting EC to migrate through the surrounding tissues.	[64]
S1P	–	Stimulates EC proliferation, migration and survival. Stimulates connective tissue growth factor (CTGF) production in ECs.	[65,66]
CTGF	–	Promotes EC growth, migration, adhesion and survival <i>in vitro</i> .	[67,68]
heparanase	–	Cleaves heparan sulfate, angiogenesis regulators binding molecule, which increases their bioavailability and facilitating their participation in blood vessel development during wound healing, tumour growth and metastasis.	[6]
ANGIOGENESIS INHIBITORS			
Factor	Platelet concentration	Mechanism of action	Ref.
angiostatin	–	Inhibits proliferation of EC <i>in vitro</i> , formation of capillary structures <i>in vitro</i> and angiogenesis <i>in vivo</i> .	[69–71]
TSP-1	31 ± 12 ng/10 ⁶ PLT	Inhibits EC proliferation and capillary tube formation. It binds CD36 on the endothelial surface and activates a signalling cascade leading to stimulation of caspase-3 and increased EC apoptosis.	[45,72–77]
PF4/CXCL4 and CXCL4L1/PF4var	12 ± 5 ng/10 ⁶ PLT	Inhibits binding of angiogenesis stimulators (e.g. VEGF, FGF) to cells.	[45,78–83]
endostatin	5.6 ± 3.0 pg/10 ⁶ PLT	Inhibits tumour growth and VEGF-induced angiogenesis, but the mechanism of its action remains unclear.	[45,84]
TIMPs (TIMP-4, TIMP-1)	TIMP-4: 120–160 pg/10 ⁶ PLT TIMP-1: < 10 pg/10 ⁶ PLT	Hinder the angiogenic process via neutralization of the activity of different MMPs.	[85]

EC – Endothelial Cell, VEGF – Vascular Endothelial Growth Factor, PDGF – Platelet-Derived Growth Factor, FGFs – Fibroblast Growth Factors, EGF – Epidermal Growth Factor, HGF – Hepatocyte Growth Factor, IGF – Insulin-Like Growth Factor, SDF-1 – Stromal Cell-derived Factor-1, MMPs – Matrix Metalloproteinases, S1P – Sphingosine-1-phosphate, CTGF – Connective Tissue Growth Factor, TSP1 – Thrombospondin-1, PF4 – Platelet Factor 4, TIMPs – Tissue Inhibitors of Metalloproteinases

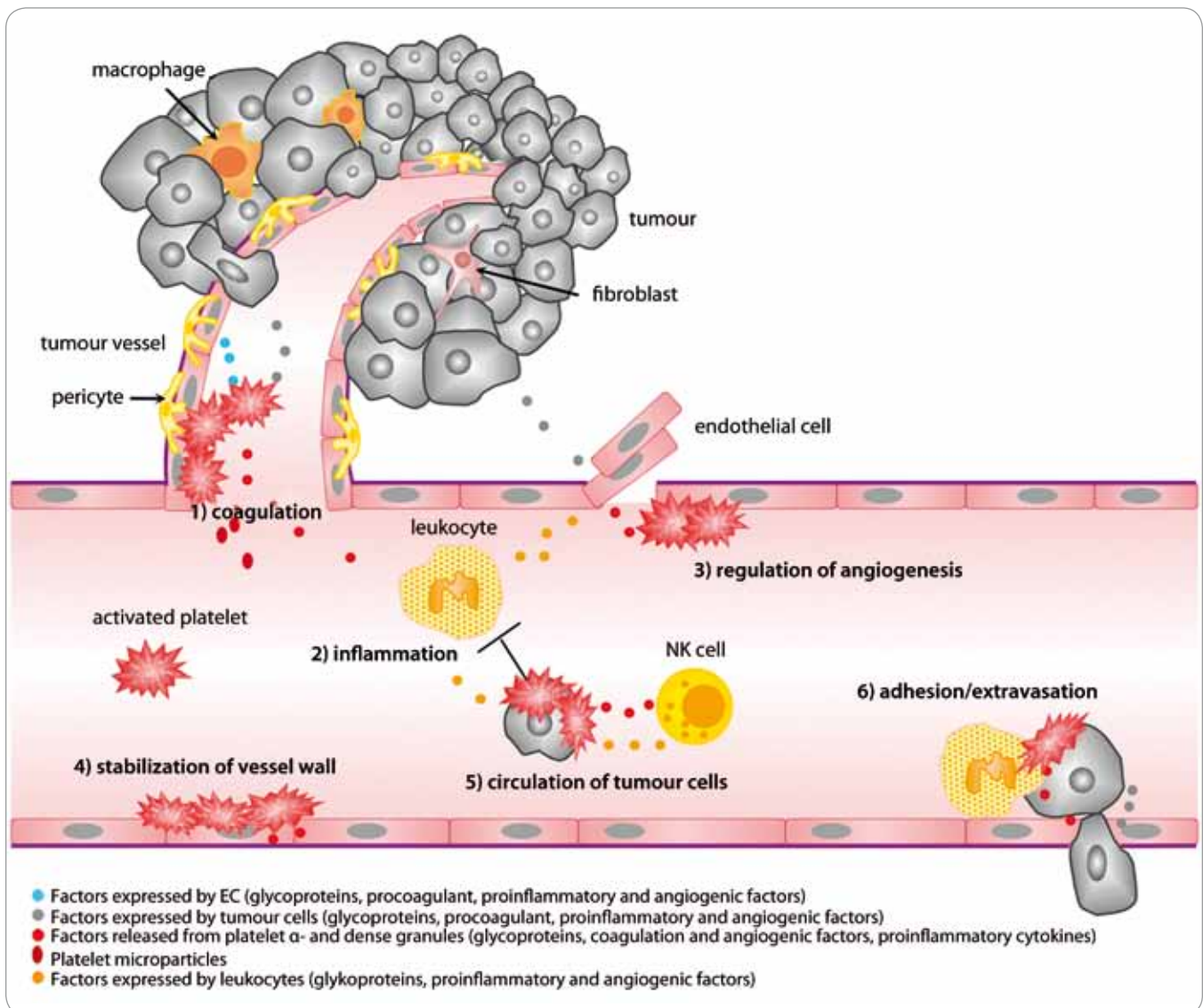


Fig. 1. Platelets contribution to the regulation of tumour angiogenesis and tumour progression. 1. Coagulation: Stimuli for platelet activation come from endothelial cells, as well as tumour stroma itself (expression of tissue factor, thrombin, ADP etc.). After activation, platelets change their shape, release PMP, α - and dense granule content and trigger coagulation cascade [8,11]. 2. Inflammation: Chemokines (IL-8, histamine etc.) released by platelets are chemotactic for leukocytes and precursor cells from bone marrow. These cells also regulate the tumour environment by release of growth and angiogenic factors [12]. 3. Angiogenesis: Platelets participate also in regulation of angiogenesis by releasing pro- and anti-angiogenic factors (VEGF, bFGF, PF-4 etc.), as well as by active sequestering of factors from the circulation [9]. 4. Stabilisation of vessel wall: Platelets stabilise the vessel wall and maintain intercellular connections by releasing factors, such as EGF, S1P, ang-1 etc., to prevent haemorrhage at the site of angiogenesis and inflammation [17]. 5. Circulation of tumour cells: Platelets adhered to tumour cells protect them from immune recognition and the cytotoxic effects of NK cell cytokines, which enables survival in the circulation and migration to distant tissue sites [7,17,18]. 6. Adhesion/extravasation: Aggregates of platelets, leukocytes and tumour cells facilitate adhesion of tumour cells to endothelium and subsequent extravasation into distant tissues. Platelets also release factors promoting cell proliferation and increasing permeability of the vessel wall (e.g. VEGF) [6,17].

detectable changes in platelet levels of angiogenesis regulators [41,93].

Are Platelets Stimulatory or Inhibitory to Tumour Growth?

While postulated many decades ago, the consequences of platelet adhesion

to activated endothelium, and their role in early tumour growth and tumour angiogenesis, has been difficult to establish. The main source of the difficulties, similar to the difficulties in establishing their role in wound healing, is the variable method of platelet concentrate pre-

paration. An additional limitation is the animal models, which do not necessarily reciprocate the complexity of platelet receptors and tissue integrins. However, through the use of genetically altered animals, *in vivo* tracking dyes, and three dimensional *in vitro* models, some of the

interactions between platelets, tumour cells, and other inflammatory cells within the tumour microenvironment are beginning to emerge (Fig. 1). The early literature can be very confusing. For example, there is convincing evidence that platelets enhance the development of metastasis [2,13–15,94–97] and primary tumour growth [2,13,15,98], but some studies advocate that the effect of platelets on primary tumour growth is inhibitory [99–101] and that the inhibition of platelet adhesion leads to promotion of metastasis [102]. Similarly, the most abundant proteins in platelets, e.g. PF4 (Tab. 1) are very potent inhibitors of tumour growth [103–109] and other very abundant platelet-associated proteins such as thrombospondin (TSP1) (Tab. 1), previously thought to be inhibitory to angiogenesis [110], may be augmenting the metastatic process under specific conditions [111–112].

One possible explanation for these very contradictory findings may be that platelets are neither inhibitors nor stimulators of tumour growth. Similar to their function in wound healing, they modulate rather than stimulate the malignant process, and the overall result of the platelet effect may depend on the balance of stimulatory and inhibitory signals within the tumour microenvironment. Depending on the reciprocal interaction between the existing host stromal cells, the oncogene-transformed tumour cells, and the recruited progenitors and inflammatory cells, the sum of these communications determines whether the outcome is growth, dormancy, or regression [94]. The final response may be less dependent on platelet numbers than on the specific content of stimulators and inhibitors of angiogenesis in the α -granules of platelets. This content of growth stimulators and inhibitors is continuously modified, a process aided by the short half-life of platelets in circulation (4–7 days in mice and 7–10 days in humans). This theory is informed by the recent finding that there is a higher organization of the opposing angiogenesis-related activities in platelets, enabling a differential release of either stimulators or inhibitors of angiogenesis [113,119]. The stimulators of angiogenesis

(e.g., VEGF and bFGF) do not reside in the same granules as the inhibitors (e.g., endostatin) [113].

A widely-held assumption is that platelets degranulate en masse upon activation, and that serum is a good reflection of their content [114–117]. This assumption, which may have hindered the understanding of the reciprocal interaction of platelets and tissues, may not be entirely correct. Angiogenesis regulators associated with α -granules of platelets, unlike the proteins of dense granules, are not indiscriminately released in response to ADP, thrombin or epinephrine [41]. Activation of human platelets with adenosine diphosphate (ADP) stimulates the release of VEGF, but not endostatin whereas, thromboxane A(2) (TXA(2)) releases endostatin but not VEGF [118]. As has been well documented in the setting of gastric ulcers, platelet responses to thrombin are also graded [119–122]. Activation of high-affinity thrombin receptor PAR1 releases stimulators such as VEGF, whereas the low affinity thrombin receptor PAR4 mainly releases inhibitors such as endostatin [121]. Similarly, the increases in acidity and temperature, which are typical in the setting of infection, inflammation, or cancer also change the sequence of release of angiogenesis regulators from platelets [123,124]. This concept may be quite intuitive: if platelets contain both stimulators and inhibitors of angiogenesis, a massive degranulation would be unlikely to provide the sustained and carefully orchestrated signals required for modulation of normal angiogenesis. It is much more advantageous if the majority of angiogenesis regulators sequestered in platelets during early cancer development remain associated with the platelet clot upon coagulation [41]. Some may even be taken up by platelets during activation [125]. This finding may provide some early insights into the mechanisms of tissue/platelet communication. Because the majority of proteins relevant to angiogenesis are retained in the α -granules of platelets, and because the organization of proteins within the α -granules is based on function [113]; the release of angiogenesis regulators, unlike the rele-

ase of ADP and serotonin from the dense granules, is selective [113,120,121], but also amenable to influences beyond the proteolytic activity of thrombin or environmental influences such as temperature or acidity. In the setting of this new knowledge, a clot, which has been thought of as a simple “plug” to prevent bleeding, now appears to be a sophisticated matrix that is rich in proteins and can regulate angiogenesis and inflammation in a locally-defined, reciprocal fashion.

Platelet-Derived Microparticles in Tumour Progression

Elevation of platelet-derived microparticles (PMP) levels accompanies a number of disorders including cancer, atherosclerosis, sepsis and diabetes [126]. The role of PMP in disease development is unknown but the composition of PMP in the plasma of patients varies considerably depending on the severity of the pathology [127]. The method of cell-cell communication may be dependent on the shedding of PMP upon platelet activation. PMP host a variety of cytokines and growth factors modulating angiogenesis and tissue regeneration. PMP have been shown to promote proliferation of endothelial cells and tubule formation [128] but also survival and proliferation of other cell types [129,130]. Recent evidence suggests that PMP, much like platelets, significantly affect tumour metastasis including modification of angiogenic responses. In gastric cancer, Kim et al showed that PMP levels are better predictors of metastasis than VEGF, IL-6, and RANTES [131]. It has been reported that PMP may serve as chemoattractants to several lung cancer cell lines, activating phosphorylation of ERK and expression of membrane type 1-matrix metalloproteinase [132]. PMP were also shown to stimulate proliferation and adhesion of cancer cells to fibrinogen and EC and enhance the adhesion and chemoinvasion of breast cancer cell lines [130]. PMP can induce secretion of MMP-2 by prostate cancer cells *in vitro*, facilitating their passage through the collagen that is a major component of extracellular matrix [133] contributing to cancer cell spread.

In summary, the systems biology of cancer is not dissimilar from that of a wound. In general, platelets have a pro-angiogenic effect in the setting of early injury, progressive tumour growth, atherogenesis or chronic inflammation, and an anti-angiogenic effect in the setting of a healing wound, dormant tumours, or receding inflammation [94]. Cancer may be thought of as “a wound that never heals” [86,87].

References

- Pearlstein E, Ambrogio C, Gasic G et al. Inhibition of the platelet-aggregating activity of two human adenocarcinomas of the colon and an anaplastic murine tumor with a specific thrombin inhibitor, dansylarginine N-(3-ethyl-1,5-pentanediy)amide. *Cancer Res* 1981; 41(11 Pt 1): 4535–4539.
- Gasic GJ, Gasic TB, Galanti N et al. Platelet-tumor-cell interactions in mice. The role of platelets in the spread of malignant disease. *Int J Cancer* 1973; 11(3): 704–718.
- Donati MB, Falanga A. Pathogenetic mechanisms of thrombosis in malignancy. *Acta Haematol* 2001; 106(1–2): 18–24.
- Pinedo HM, Verheul HM, D'Amato RJ et al. Involvement of platelets in tumour angiogenesis? *Lancet* 1998; 352(9142): 1775–1777.
- Verheul HM, Pinedo HM. Tumor Growth: A Putative Role for Platelets? *Oncologist* 1998; 3(2): 11.
- Vlodavsky I, Eldor A, Haimovitz-Friedman A et al. Expression of heparanase by platelets and circulating cells of the immune system: possible involvement in diapedesis and extravasation. *Invasion Metastasis* 1992; 12(2): 112–127.
- Zhou J, Sargiannidou I, Tuszyński GP. The role of adhesive proteins in the hematogenous spread of cancer. *In Vivo* 2000; 14(1): 199–208.
- Milsom C, Rak J. Tissue factor and cancer. *Pathophysiol Haemost Thromb* 2008; 36(3–4): 160–176.
- Sierko E, Wojtukiewicz MZ. Platelets and angiogenesis in malignancy. *Semin Thromb Hemos* 2004; 30(1): 95–108.
- Sierko E, Wojtukiewicz MZ. Inhibition of platelet function: does it offer a chance of better cancer progression control? *Semin Thromb Hemos* 2007; 33(7): 712–721.
- ten Cate H, Falanga A. Overview of the postulated mechanisms linking cancer and thrombosis. *Pathophysiol Haemost Thromb* 2008; 36(3–4): 122–130.
- Weyrich AS, Lindemann S, Zimmerman GA. The evolving role of platelets in inflammation. *J Thromb Haemost* 2003; 1(9): 1897–1905.
- Gasic GJ, Gasic TB, Stewart CC. Antimetastatic effects associated with platelet reduction. *P Natl Acad Sci USA* 1968; 61(1): 46–52.
- Borsig L. The role of platelet activation in tumor metastasis. *Expert Rev Anticancer Ther* 2008; 8(8): 1247–1255.
- Gasic GJ. Role of plasma, platelets, and endothelial cells in tumor metastasis. *Cancer Metast Rev* 1984; 3(2): 99–114.
- Borsig L. Antimetastatic activities of modified heparins: selectin inhibition by heparin attenuates metastasis. *Semin Thromb Hemos* 2007; 33(5): 540–546.
- Erpenbeck L, Schon MP. Deadly allies: the fatal interplay between platelets and metastasizing cancer cells. *Blood* 2010; 115(17): 3427–3436.
- Palumbo JS, Talmage KE, Massari JV et al. Platelets and fibrin(ogen) increase metastatic potential by impeding natural killer cell-mediated elimination of tumor cells. *Blood* 2005; 105(1): 178–185.
- Connolly GC, Khorana AA. Risk stratification for cancer-associated venous thromboembolism. *Best Pract Res Clin Haematol* 2009; 22(1): 35–47.
- Khorana AA, Connolly GC. Assessing risk of venous thromboembolism in the patient with cancer. *J Clin Oncol* 2009; 27(29): 4839–4847.
- Lee AY, Levine MN. Venous thromboembolism and cancer: risks and outcomes. *Circulation* 2003; 107(23 Suppl 1): 117–121.
- Khorana AA. Risk assessment for cancer-associated thrombosis: what is the best approach? *Thromb Res* 2012; 129(Suppl 1): S10–S15.
- Berthold F, Sahin K, Hero B et al. The current contribution of molecular factors to risk estimation in neuroblastoma patients. *Eur J Cancer* 1997; 33(12): 2092–2097.
- Engan T, Hannisdal E. Blood analyses as prognostic factors in primary lung cancer. *Acta Oncol* 1990; 29(2): 151–154.
- Lopes A, Daras V, Cross PA et al. Thrombocytosis as a prognostic factor in women with cervical cancer. *Cancer* 1994; 74(1): 90–92.
- Rosenthal MC, Niemetz J, Wisch N. Hemorrhage and thromboses associated with neoplastic disorders. *J Chronic Dis* 1963; 16: 667–675.
- Trousseau A. *Phlegmatia alba dolens*. Paris: JB Baillere et Fils 1865.
- Casu B, Vlodavsky I, Sanderson RD. Non-anticoagulant heparins and inhibition of cancer. *Pathophysiol Haemost Thromb* 2008; 36(3–4): 195–203.
- Gerotziapas GT, Papageorgiou C, Hatmi M et al. Clinical studies with anticoagulants to improve survival in cancer patients. *Pathophysiol Haemost Thromb* 2008; 36(3–4): 204–211.
- Kakkar AK. An expanding role for antithrombotic therapy in cancer patients. *Cancer Treat Rev* 2003; 29(Suppl 2): 23–26.
- Kakkar AK, Levine MN, Kadziola Z et al. Low molecular weight heparin, therapy with dalteparin, and survival in advanced cancer: the fragmin advanced malignancy outcome study (FAMOUS). *J Clin Oncol* 2004; 22(10): 1944–1948.
- Kakkar AK, Macbeth F. Antithrombotic therapy and survival in patients with malignant disease. *Br J Cancer* 2010; 102(Suppl 1): S24–S29.
- Kakkar AK, Williamson RC. Thromboprophylaxis in the cancer patient. *Haemostasis* 1998; 28(Suppl 3): 61–65.
- Petralia GA, Lemoine NR, Kakkar AK. Mechanisms of disease: the impact of antithrombotic therapy in cancer patients. *Nat Clin Pract Oncol* 2005; 2(7): 356–363.
- Thodiyil P, Kakkar AK. Can low-molecular-weight heparins improve outcome in patients with cancer? *Cancer Treat Rev* 2002; 28(3): 151–155.
- Lyman GH, Khorana AA, Falanga A et al. American Society of Clinical Oncology guideline: recommendations for venous thromboembolism prophylaxis and treatment in patients with cancer. *J Clin Oncol* 2007; 25(34): 5490–5505.
- Gimbrone MA Jr, Aster RH, Cotran RS et al. Preservation of vascular integrity in organs perfused in vitro with a platelet-rich medium. *Nature* 1969; 222(5188): 33–36.
- Gore I, Takada M, Austin J. Ultrastructural basis of experimental thrombocytopenic purpura. *Arch Pathol* 1970; 90(3): 197–205.
- Kitchens CS, Weiss L. Ultrastructural changes of endothelium associated with thrombocytopenia. *Blood* 1975; 46(4): 567–578.
- Saba SR, Mason RG. Effects of platelets and certain platelet components on growth of cultured human endothelial cells. *Thromb Res* 1975; 7(5): 807–812.
- Klement GL, Yip TT, Cassiola F et al. Platelets actively sequester angiogenesis regulators. *Blood* 2009; 113(12): 2835–2842.
- Klement G, Shai E, Varon D. The role of platelets in angiogenesis. In: Michelson A, editor. *Platelets*. 3rd ed. San Diego, CA: Elsevier/Academic Press 2012.
- Semple JW, Italiano JE Jr, Freedman J. Platelets and the immune continuum. *Nat Rev Immunol* 2011; 11(4): 264–274.
- Mohle R, Green D, Moore MA et al. Constitutive production and thrombin-induced release of vascular endothelial growth factor by human megakaryocytes and platelets. *P Natl Acad Sci USA* 1997; 94(2): 663–668.
- Peterson JE, Zurakowski D, Italiano JE et al. Normal ranges of angiogenesis regulatory proteins in human platelets. *Am J Hematol* 2010; 85(7): 487–493.
- Gerhardt H. VEGF and endothelial guidance in angiogenic sprouting. *Organogenesis* 2008; 4(4): 241–246.
- Mannaioni PF, Di Bello MG, Masini E. Platelets and inflammation: role of platelet-derived growth factor, adhesion molecules and histamine. *Inflamm Res* 1997; 46(1): 4–18.
- Heldin CH. Simultaneous induction of stimulatory and inhibitory signals by PDGF. *FEBS Lett* 1997; 410(1): 17–21.
- Brunner G, Nguyen H, Gabrilove J et al. Basic fibroblast growth factor expression in human bone marrow and peripheral blood cells. *Blood* 1993; 81(3): 631–638.
- Nakamura T, Kasai K, Banba N et al. Release of human epidermal growth factor from platelets in accordance with aggregation in vitro. *Endocrinol Jpn* 1989; 36(1): 23–28.
- Vilorio-Petit A, Crombet T, Jothy S et al. Acquired resistance to the antitumor effect of epidermal growth factor receptor-blocking antibodies in vivo: a role for altered tumor angiogenesis. *Cancer Res* 2001; 61(13): 5090–5101.
- Lee YM, Bae HM, Lee OH. Synergistic induction of in vivo angiogenesis by the combination of insulin-like growth factor-II and epidermal growth factor. *Oncol Rep* 2004; 12(4): 843–848.
- Nakamura Y, Morishita R, Higaki J et al. Expression of local hepatocyte growth factor system in vascular tissues. *Biochem Biophys Res Commun* 1995; 215(2): 483–488.
- Shima N, Itagaki Y, Nagao M et al. A fibroblast-derived tumor cytotoxic factor/F-TGF (hepatocyte growth factor/HGF) has multiple functions in vitro. *Cell Biol Int Rep* 1991; 15(5): 397–408.
- Tomita N, Morishita R, Taniyama Y et al. Angiogenic property of hepatocyte growth factor is dependent on upregulation of essential transcription factor for angiogenesis, ets-1. *Circulation* 2003; 107(10): 1411–1417.
- Karey KP, Sirbasku DA. Human platelet-derived mitogens. II: subcellular localization of insulin like growth factor I to the alpha-granule and release in response to thrombin. *Blood* 1989; 74: 1093–1100.
- Chan K, Spencer EM. Megakaryocytes endocytose insulin-like growth factor (IGF) I and IGF-binding protein-3: a novel mechanism directing them into alpha granules of platelets. *Endocrinol* 1998; 139: 559–565.
- Shigematsu S, Yamauchi K, Nakajima K et al. IGF-1 regulates migration and angiogenesis of human endothelial cells. *Endocr J* 1999; 46(Suppl): S59–S52.
- Nicosia RF, Nicosia SV, Smith M. Vascular endothelial growth factor, platelet-derived growth factor, and insulin-like growth factor-1 promote rat aortic angiogenesis in vitro. *Am J Pathol* 1994; 145(5): 1023–1029.
- Caine GJ, Lip GY, Blann AD. Platelet-derived VEGF, Flt-1, angiopoietin-1 and P-selectin in breast and prostate cancer: further evidence for a role of platelets in tumour angiogenesis. *Ann Med* 2004; 36(4): 273–277.
- Li JJ, Huang YQ, Basch R et al. Thrombin induces the release of angiopoietin-1 from platelets. *Thromb Haemost* 2001; 85(2): 204–206.
- Moore MA, Hattori K, Heissig B. Mobilization of endothelial and hematopoietic stem and progenitor cells by adenovector-mediated elevation of serum levels of SDF-1, VEGF, and angiopoietin-1. *Ann NY Acad Sci* 2001; 938: 36–45.
- Deregibus MC, Buttiglieri S, Russo S. CD40-dependent activation of phosphatidylinositol 3-kinase/Akt pathway

- mediates endothelial cell survival and in vitro angiogenesis. *J Biol Chem* 2003; 278(20): 18008–18014.
64. Mott JD, Werb Z. Regulation of matrix biology by matrix metalloproteinases. *Curr Opin Cell Biol* 2004; 16(5): 558–564.
65. Brindley DN. Lipid phosphate phosphatases and related proteins: signaling functions in development, cell division, and cancer. *J Cell Biochem* 2004; 92(5): 900–912.
66. Markiewicz M, Nakerakanti SS, Kapanadze B. Connective tissue growth factor (CTGF/CCN2) mediates angiogenic effect of S1P in human dermal microvascular endothelial cells. *Microcirculation* 2011; 18(1): 1–11.
67. Cicha I, Yilmaz A, Suzuki Y. Connective tissue growth factor is released from platelets under high shear stress and is differentially expressed in endothelium along atherosclerotic plaques. *Clinical Hemorheol Micro* 2006; 35(1–2): 203–206.
68. Yang F, Tuxhorn JA, Ressler SJ et al. Stromal expression of connective tissue growth factor promotes angiogenesis and prostate cancer tumorigenesis. *Cancer Res* 2005; 65: 8887–8895.
69. Jurasz P, Alonso D, Castro-Blanco S et al. Generation and role of angiostatin in human platelets. *Blood* 2003; 102: 3217–3223.
70. O'Reilly MS, Holmgren L, Shing Y et al. Angiostatin: a novel angiogenesis inhibitor that mediates the suppression of metastases by a Lewis lung carcinoma. *Cell* 1994; 79: 315–328.
71. O'Reilly MS, Holmgren L, Chen C et al. Angiostatin induces and sustains dormancy of human primary tumors in mice. *Nat Med* 1996; 2: 689–692.
72. Jaffe EA, Leung LL, Nachman RL et al. Thrombospondin is the endogenous lectin of human platelets. *Nature* 1982; 295(5846): 246–248.
73. Jiménez B, Volpert OV, Crawford SE et al. Signals leading to apoptosis-dependent inhibition of neovascularization by thrombospondin-1. *Nat Med* 2000; 6(1): 41–48.
74. Lawler PR, Lawler J. Molecular basis for the regulation of angiogenesis by thrombospondin-1 and -2. *Cold Spring Harb Perspect Med* 2012; 2(5): a006627.
75. Gupta K, Gupta P, Wild R et al. Binding and displacement of vascular endothelial growth factor (VEGF) by thrombospondin: effect on human microvascular endothelial cell proliferation and angiogenesis. *Angiogenesis* 1999; 3(2): 147–158.
76. Dardik R, Solomon A, Loscalzo J et al. Novel proangiogenic effect of factor XIII associated with suppression of thrombospondin 1 expression. *Arterioscler Thromb Vasc Biol* 2003; 23(8): 1472–1477.
77. Dawson DW, Volpert OV, Pearce SF et al. Three distinct D-amino acid substitutions confer potent antiangiogenic activity on an inactive peptide derived from a thrombospondin-1 type 1 repeat. *Mol Pharmacol* 1999; 55(2): 332–338.
78. Kowalska MA, Rauova L, Poncz M. Role of the platelet chemokine platelet factor 4 (PF4) in hemostasis and thrombosis. *Thromb Res* 2010; 125(4): 292–296.
79. Bikfalvi A. Platelet factor 4: an inhibitor of angiogenesis. *Semin Thromb Hemost* 2004; 30(3): 379–385.
80. Chadderton NS, Stringer SE. Interaction of platelet factor 4 with fibroblast growth factor 2 is stabilised by heparan sulphate. *Int J Biochem Cell Biol* 2003; 35(7): 1052–1055.
81. Kim SH, Kiick KL. Heparin-mimetic sulfated peptides with modulated affinities for heparin-binding peptides and growth factors. *Peptides* 2007; 28(11): 2125–2136.
82. Hagedorn M, Zilberberg L, Wilting J et al. Domain swapping in a COOH-terminal fragment of platelet factor 4 generates potent angiogenesis inhibitors. *Cancer Res* 2002; 62(23): 6884–6890.
83. Vandercappellen J, Liekens S, Bronckaers A et al. The COOH-terminal peptide of platelet factor-4 variant (CXCL4L1/PF-4var47-70) strongly inhibits angiogenesis and suppresses B16 melanoma growth in vivo. *Mol Cancer Res* 2010; 8(3): 322–334.
84. Xu HL, Tan HN, Wang FS et al. Research advances of endostatin and its short internal fragments. *Curr Protein Pept Sci* 2008; 9(3): 275–283.
85. Radomski A, Jurasz P, Sanders EJ et al. Identification, regulation and role of tissue inhibitor of metalloproteinases-4 (TIMP-4) in human platelets. *Br J Pharmacol* 2002; 137(8): 1330–1338.
86. Dvorak HF. Tumors: wounds that do not heal. Similarities between tumor stroma generation and wound healing. *N Engl J Med* 1986; 315(26): 1650–1659.
87. Dvorak HF, Harvey VS, Estrella P et al. Fibrin containing gels induce angiogenesis. Implications for tumor stroma generation and wound healing. *Lab Invest* 1987; 57(6): 673–686.
88. Nagy JA, Brown LF, Senger DR et al. Pathogenesis of tumor stroma generation: a critical role for leaky blood vessels and fibrin deposition. *Biochim Biophys Acta* 1989; 948(3): 305–326.
89. Rak J, Yu JL, Klement G et al. Oncogenes and angiogenesis: signaling three-dimensional tumor growth. *J Invest Dermatol Symp Proc* 2000; 5(1): 24–33.
90. Rak J, Yu JL, Kerbel RS et al. What do oncogenic mutations have to do with angiogenesis/vascular dependence of tumors? *Cancer Res* 2002; 62(7): 1931–1934.
91. Rak J, Klement G. Impact of oncogenes and tumor suppressor genes on deregulation of hemostasis and angiogenesis in cancer. *Cancer Metastasis Rev* 2000; 19(1–2): 93–96.
92. Rak J, Filmus J, Finkenzeller G et al. Oncogenes as inducers of tumor angiogenesis. *Cancer Metastasis Rev* 1995; 14(4): 263–277.
93. Cervi D, Yip TT, Bhattacharya N et al. Platelet-associated PF-4 as a biomarker of early tumor growth. *Blood* 2008; 111(3): 1201–1207.
94. Almog NK, Klement GL. Platelet Proteome and Tumor Dormancy: Can Platelets Content Serve as Predictive Biomarkers for Exit of Tumors from Dormancy? *Cancers* 2010; 2(2): 842–858.
95. Ellis LM, Fidler IJ. Angiogenesis and metastasis. *Eur J Cancer* 1996; 32A(14): 2451–2460.
96. Mehta P. Potential role of platelets in the pathogenesis of tumor metastasis. *Blood* 1984; 63(1): 55–63.
97. Yahalom J, Eldor A, Biran S et al. Platelet-tumor cell interaction with the subendothelial extracellular matrix: relationship to cancer metastasis. *Radiother Oncol* 1985; 3(3): 211–225.
98. Nierodzik ML, Karpatkin S. Thrombin induces tumor growth, metastasis, and angiogenesis: Evidence for a thrombin-regulated dormant tumor phenotype. *Cancer Cell* 2006; 10(5): 355–362.
99. Daly ME, Makris A, Reed M et al. Hemostatic regulators of tumor angiogenesis: a source of antiangiogenic agents for cancer treatment? *J Natl Cancer Inst* 2003; 95(22): 1660–1673.
100. Ibele G, Kay N, Johnson G et al. Human platelets exert cytotoxic effects on tumor cells. *Blood* 1985; 65(5): 1252–1255.
101. Wang Y, Zhang H. Platelet-induced inhibition of tumor cell growth. *Thromb Res* 2008; 123(2): 324–330.
102. Erpenbeck L, Nieswandt B, Schon M et al. Inhibition of platelet GPIIb/IIIa and promotion of melanoma metastasis. *J Invest Dermatol* 2010; 130(2): 576–586.
103. Bikfalvi A. Platelet factor 4: an inhibitor of angiogenesis. *Semin Thromb Hemost* 2004; 30(3): 379–385.
104. Kolber DL, Knisely TL, Maione TE. Inhibition of development of murine melanoma lung metastases by systemic administration of recombinant platelet factor 4. *J Natl Cancer Inst* 1995; 87(4): 304–309.
105. Vandercappellen J, Van Damme J, Struyf S. The role of the CXCL chemokines platelet factor-4 (CXCL4/PF-4) and its variant (CXCL4L1/PF-4var) in inflammation, angiogenesis and cancer. *Cytokine Growth Factor Rev* 2011; 22(1): 1–18.
106. Vandercappellen J, Liekens S, Bronckaers A et al. The COOH-terminal peptide of platelet factor-4 variant (CXCL4L1/PF-4var47-70) strongly inhibits angiogenesis and suppresses B16 melanoma growth in vivo. *Mol Cancer Res* 2010; 8(3): 322–334.
107. Hagedorn M, Zilberberg L, Wilting J et al. Domain swapping in a COOH-terminal fragment of platelet factor 4 generates potent angiogenesis inhibitors. *Cancer Res* 2002; 62(23): 6884–6890.
108. Maione TE, Gray GS, Petro J et al. Inhibition of angiogenesis by recombinant human platelet factor-4 and related peptides. *Science* 1990; 247(4938): 77–79.
109. Yamaguchi K, Ogawa K, Katsube T et al. Platelet factor 4 gene transfection into tumor cells inhibits angiogenesis, tumor growth and metastasis. *Anticancer Res* 2005; 25(2A): 847–851.
110. Tuszynski GP, Nicosia RF. The role of thrombospondin-1 in tumor progression and angiogenesis. *Bioessays* 1996; 18(1): 71–76.
111. Tuszynski GP, Gasic TB, Rothman VL et al. Thrombospondin, a potentiator of tumor cell metastasis. *Cancer Res* 1987; 47(15): 4130–4133.
112. Walz DA. Thrombospondin as a mediator of cancer cell adhesion in metastasis. *Cancer Metastasis Rev* 1992; 11(3–4): 313–324.
113. Italiano JE Jr, Richardson JL, Patel-Hett S et al. Angiogenesis is regulated by a novel mechanism: pro- and antiangiogenic proteins are organized into separate platelet alpha granules and differentially released. *Blood* 2008; 111(3): 1227–1233.
114. Benoy I, Salgado R, Colpaert C et al. Serum interleukin 6, plasma VEGF, serum VEGF, and VEGF platelet load in breast cancer patients. *Clin Breast Cancer* 2002; 2(4): 311–315.
115. Caine GJ, Lip GY, Blann AD. Platelet-derived VEGF, Flt-1, angiopoietin-1 and P-selectin in breast and prostate cancer: further evidence for a role of platelets in tumour angiogenesis. *Ann Med* 2004; 36(4): 273–277.
116. Lee JK, Hong YJ, Han CJ et al. Clinical usefulness of serum and plasma vascular endothelial growth factor in cancer patients: which is the optimal specimen? *Int J Oncol* 2000; 17(1): 149–152.
117. Werther K, Christensen IJ, Nielsen HJ. Prognostic impact of matched preoperative plasma and serum VEGF in patients with primary colorectal carcinoma. *Br J Cancer* 2002; 86(3): 417–423.
118. Battinelli EM, Markens BA, Italiano JE Jr. Release of angiogenesis regulatory proteins from platelet alpha granules: modulation of physiologic and pathologic angiogenesis. *Blood* 2011; 118(5): 1359–1369.
119. Ma L, Elliott SN, Cirino G et al. Platelets modulate gastric ulcer healing: role of endostatin and vascular endothelial growth factor release. *Proc Natl Acad Sci U S A* 2001; 98(11): 6470–6475.
120. Ma L, Hollenberg MD, Wallace JL. Thrombin-induced platelet endostatin release is blocked by a proteinase-activated receptor-4 (PAR4) antagonist. *Br J Pharmacology* 2001; 134(4): 701–704.
121. Ma L, Perini R, McKnight W et al. Proteinase-activated receptors 1 and 4 counter-regulate endostatin and VEGF release from human platelets. *Proc Natl Acad Sci U S A* 2005; 102(1): 216–220.
122. Perini R, Wallace JL, Ma L. Roles of platelets and proteinase-activated receptors in gastric ulcer healing. *Dig Dis Sci* 2005; 50 (Suppl 1): S12–S15.
123. Etulain J, Laponni MJ, Patrucchi SJ et al. Hyperthermia inhibits platelet hemostatic functions and selectively regulates the release of alpha-granule proteins. *J Thromb Haemostasis* 2011; 9(8): 1562–1571.
124. Etulain J, Negrotto S, Carestia A et al. Acidosis downregulates platelet haemostatic functions and pro-

- motes neutrophil proinflammatory responses mediated by platelets. *J Thromb Haemost* 2011; 107(1): 99–110.
- 125.** Akerblom B, Lindahl TL, Larsson A. ADP activation induces bFGF binding to platelets in vitro. *Ups J Med Sci* 2002; 107(3): 165–171.
- 126.** Simak J, Gelderman MP. Cell membrane microparticles in blood and blood products: potentially pathogenic agents and diagnostic markers. *Transfus Med Rev* 2006; 20(1): 1–26.
- 127.** Helley D, Banu E, Bouziane A et al. Platelet microparticles: a potential predictive factor of survival in hormone-refractory prostate cancer patients treated with docetaxel-based chemotherapy. *Eur Urol* 2009; 56(3): 479–484.
- 128.** Kim HK, Song KS, Chung JH et al. Platelet microparticles induce angiogenesis in vitro. *Br J Haematol* 2004; 3: 376–384.
- 129.** Hayon Y, Dashevsky O, Shai E et al. Platelet microparticles promote neural stem cell proliferation, survival and differentiation. *J Mol Neurosci* 2012; 47(3): 659–665.
- 130.** Janowska-Wieczorek A, Marquez-Curtis LA, Wysoczynski M et al. Enhancing effect of platelet-derived microvesicles on the invasive potential of breast cancer cells. *Transfusion* 2006; 46(7): 1199–1209.
- 131.** Kim HK, Song KS, Park YS et al. Elevated levels of circulating platelet microparticles, VEGF, IL-6 and RANTES in patients with gastric cancer: possible role of a metastasis predictor. *Eur J Cancer* 2003; 39(2): 184–191.
- 132.** Janowska-Wieczorek A, Wysoczynski M, Kijowski J et al. Microvesicles derived from activated platelets induce metastasis and angiogenesis in lung cancer. *Int J Cancer* 2005; 113(5): 752–760.
- 133.** Dashevsky O, Varon D, Brill A. Platelet-derived microparticles promote invasiveness of prostate cancer cells via upregulation of MMP-2 production. *Int J Cancer* 2009; 124(8): 1773–1777.